

Original paper

Title: Toxicological evaluation of *Aloe barbadensis* root extracts in Wistar rats: Haematology and Lipid profile.

Running title: Effect of *Aloe barbadensis* root extract on Wistar rat's haematology and lipid profile.

ABSTRACT

A. barbadensis is a folkloric medicinal plant used for decades to treat several ailments such as intestinal ulcers, gynaecological problems, wound healing, ringworm and eczema. Other uses of *A. barbadensis* include impotence, low libido, appetite disorder, emmenagogue, pile, asthma, cough and jaundice. This study was aimed at determining the safety of the ethanol extract of *Aloe barbadensis* root using haematological, and lipid parameters. *A. barbadensis* root extract (100, 200 and 400 mg/kg) and control (distilled water-0.2mL/kg) were administered to sixty male Wistar rats (150-270 g body weights) for 14 days. The haematological parameters were determined using the collected whole blood and lipid profile assessed using the serum. The oral administration of the extract on red blood cells and white blood cells, as well as other haematological indices (haemoglobin, Platelets crit, mean corpuscular volume, mean corpuscular haemoglobin, mean corpuscular haemoglobin concentration, granulocytes, lymphocytes, Platelets crit and monocytes), were significantly ($p < 0.05$) not altered. A non-significant ($p > 0.05$) effect of the extract on high-density lipoproteins (HDL) and low-density lipoproteins (LDL) was observed in the serum of the male rats. The results indicate that within the doses used in the study, the ethanol extract of *A. barbadensis* root is relatively non-toxic with no significant

localized toxicity. However, the root of *A. barbadensis* should be cautiously used because of its selective effect on some lipid parameters in the male rats.

Keywords: *Aloe barbadensis*; toxicity; ethanol extract; hematological parameters; lipid profile.

INTRODUCTION

Aloe barbadensis Mill. commonly called aloe vera, belongs to the family Xanthorrhoeaceae (APG, 2009). *A. barbadensis* is a succulent herb growing up to 1-4 feet tall with a short stem and white spots on the light green leaves. The leaves (20-50 cm long), 3-5 cm wide at the base, tapering to a pointed tip. *Aloe vera* possesses a shallow root (Ryckowski, 2020) with a rhizome-root system (Rajeswari et al. 2012). The folk use the leaf juice to treat intestinal ulcers, catarrh and gynaecological problems (Idu et al. 2014). Similarly, fresh leaf juice is taken orally to treat stomach ulcers and heal wounds (Ross, 1999). Aloe vera leaf can be used to manage skin irritations such as ringworm and eczema when passed over low heat. The root of *A. barbadensis*, according to Adodo (2014), can be used to treat constipation and impotence. It is also used as a purgative, appetite-stimulant, emmenagogue and for managing colds, piles, asthma, cough and jaundice in ayurvedic formulation (Joseph and Raj, 2010). The root of *A. barbadensis* is popularly used to treat low libido, a common practice amongst the Ifa Nkari people of Akwa Ibom State, Nigeria (Erhabor et al. 2013).

It is important to note that *Aloe vera*, like other medicinal plants, deserves to be screened for safety. This becomes imperative following the immense traditional uses associated with different plant parts. Though, it is commonly believed that herbs are safe by Traditional medicine practitioners and other users of medicinal plants. This folk belief underscores the perception of the plant's natural origin (Afolayan and Yakubu, 2009). Despite this general belief, medicinal

plants possess active ingredients capable of having any form of detrimental effects on man and animals. This harmful effect can be on the cell (cytotoxicity), liver (hepatotoxicity), kidney (nephrotoxicity), blood (heamatotoxicity) or lipid (lipotoxicity).

Nevertheless, these injuries' occurrences depend on the number of chemicals absorbed (Betram, 1998). It is valid that appropriate scientific investigation of any medicinal plant's beneficial and harmful effects should be done (Idu et al. 2006). The toxic consequence of a drug in man has been reported to be akin to that of certain animals. Therefore, animal models are used in toxicological studies (Range et al. 1995). Validating any chemical substance or medicinal plant's toxicity has helped determine the upper limits of effective therapy (Sofowora, 1993).

Furthermore, the dearth of toxicological information on the root of *A. barbadensis* prodded this investigation. In this pre-clinical study, the toxicity evaluation of *A. barbadensis* root was limited to male Wistar rats' haematological and lipid profiles. The folkloric use of *A. barbadensis* root as an aphrodisiac in Nigeria (Erhabor and Idu, 2017) necessitated the use of male Wistar rats. Thus, this study was carried out to provide information on *A. barbadensis* root extract's toxicity in male rats.

MATERIALS AND METHODS

Collection and extraction of plant material

The fresh roots of *A. barbadensis* were collected in Okene settlement, Kogi State, Nigeria. The plant was identified by Mr. G. Ighanesebhor, Herbarium Unit, Obafemi Awolowo University, Ile-Ife, Nigeria, with voucher number IFE17004, where it was deposited. Initially, the roots were rinsed in running water and placed on laboratory tables to dry at room

temperature. The roots were further dried in an oven set at 40 °C for 10 minutes before grinding to powder. The fine powdered plant material (2kg) was extracted with 5L of ethanol using a soxhlet extractor. The extract was concentrated to dryness using a water bath (HH-S Water Bath; Searchtech Instruments) set at an average temperature of 50 °C.

Animal grouping and administration of the extract

Sixty (60) healthy male Wistar rats (150-270 g body weight) were obtained from the animal breeding house of the Department of Anatomy, Faculty of Basic Medical Sciences, University of Benin, Nigeria, for this study. The animals were in the ventilated animal house of the Department of Animal and Environmental Biology, Faculty of Life Sciences, University of Benin, Nigeria, for acclimatization with optimum conditions (temperature, 25°C; photoperiod, 12 hours of natural light and 12 hours of dark). The animals had unfettered access to water and standard commercial pellets. The ethical committee on experimental animal use and care of the Faculty of Life Sciences, University of Benin, Nigeria, reviewed and approved the protocol (LSC15101).

The 60 male rats were randomly placed into four groups (groups A, B, C and D) of 15 animals each and given treatment orally. Group A was administered the diluent (2 mL of distilled water), while groups B, C and D were given 100, 200 and 400 mg/kg body weight of *A. barbadensis* root extract using an orogastric tube. Five rats from all the groups were sacrificed after 1¼ hours of administering the respective doses of the extract on days 1, 7 and 14. The animals were handled following the international guiding principles for biomedical research involving animals as outlined by the Council for International Organization of Medical Sciences and the International Council for Laboratory Animal Science (CIOMS and ICLAS, 2012).

Preparation of serum

The modified procedure of Yakubu et al. (2005), was adopted in the preparation of the serum. Blood was collected from the Wistar rats on days 1, 7 and 14. Briefly, the rats' abdominal cavity was cut open under chloroform anesthesia to expose the internal organs with sterile forceps and scissors. After that, blood was collected through cardiac puncture using a 5 mL syringe and needle per animal into properly labelled clean non-coagulant sample bottles. The sample bottles were left at room temperature for 10 minutes to clot. An aliquot (2 mL) of the blood was collected into ethylenediaminetetraacetic acid (EDTA) sample bottles for the haematological analysis. The bottles were centrifuged at 3000 rpm for 10 minutes using a laboratory centrifuge. The collected sera were aspirated with Pasteur pipettes into clean, dry sample plain bottles and used within 12 hours of preparation for the lipid assays.

Haematological studies

The automated Sysmex KX-21 haematology analyzer (Sysmex Corporation, Kobe, Japan) was used to determine the number of red blood cells (RBC), haemoglobin (HGB), haematocrit (HCT), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), and mean corpuscular haemoglobin concentration (MCHC). Other haematological parameters assayed include red cell distribution width (RCDW), white blood cell (WBC), monocytes (MO), lymphocytes (LY), and platelets (PLT). Platelets crit (PCT), platelet density width (PDW), mean platelet volume (MPV) and granulocytes (GR).

Determination of lipid profile

The lipid profile of the serum was assayed by evaluating the total cholesterol, triglyceride (TRIG), high-density lipoproteins (HDL) and low-density lipoproteins (LDL). The total

cholesterol was determined using Trinder's earlier protocol (1969), while the amount of TRIG was done following Tietz's procedure (Tietz, 1990). The amount of high-density lipoproteins (HDL) and low-density lipoproteins (LDL) in serum was assessed by adopting the method described in the manufacturer's kits (Randox Laboratories Limited, UK).

Analysis of Data

Analyzed data were presented as mean \pm SEM of the appropriate replicates. To compare means of different groups, One Way ANOVA was utilized and Duncan, multiple range tests done, to ascertain the differences among various means and the interaction between the variables using SPSS 15.0 computer software package. Differences at $P < 0.05$ or $P < 0.01$ were considered statistically significant.

RESULTS

Effect of *A. barbadensis* on Haematological parameters

Table 1 showed that most of the haematological parameters were not significantly affected by the extract's administration at the tested doses. The analyzed blood indices (HGB- haemoglobin, RBC-red blood cells count, MCH-mean corpuscular haemoglobin, RDW- red density width, GR- granulocytes count, LY(%)- lymphocytes, MO(%)- monocytes, PCT(%)- platelet crit, PDW(%)-platelet density width granulocyte) were not significantly affected by the administration of the extract on the respective days of treatment and across the days of testing too.

Table 1. Effect of administration of ethanol extract of *A. barbadensis* on haematological parameters of male rats

		Control(DW)	100mg/kg	200mg/kg	400mg/kg	P-value
GR (%)	Day 1	12±4.06	11.52±2.93	32.08±14.37	22.84±8.02	P>0.05
	Day 7	17.8±9.59	6.74±0.98	27.7±8.45	13.58±5.08	P>0.05
	Day 14	10.02±5.3	9.36±4.96	6.48±3.24	8.4±3.78	P>0.05
	P-Value	NS	NS	NS	NS	
HCT	Day 1	37.9±3.41	63.62±22.07	35.14±3.67	34.38 [#] ±5.18	P>0.05
	Day 7	45.88±4.24	50.46±2.78	46.88±2.82	57.66 ^{##} ±7.87	P>0.05
	Day 14	40.66±0.9	40.6±1.75	34.28±4.38	38.7 [#] ±4.55	P>0.05
	P-Value	NS	NS	NS	*P<0.05	
HGB	Day 1	13.3±1.19	12.24±0.93	12.46±0.76	12.8±1.23	P>0.05
	Day 7	14.52±1.19	15±0.72	13.98±0.73	14.74±0.44	P>0.05
	Day 14	14.44±0.39	14.26±0.57	11.54±1.64	13.28±1.36	P>0.05
	P-Value	NS	NS	NS	NS	
LY%	Day 1	79.48±6.53	80.44±5.36	73.4±9.22	66.78±10.85	P>0.05
	Day 7	76.44±12.3	88.5±1.63	59.52±11.71	78.44±8.33	P>0.05
	Day 14	80.38±10.33	81.4±10.39	86.82±7.04	84.74±7.77	P>0.05
	P-Value	NS	NS	NS	NS	
MCH	Day 1	19.42±0.47	13.9±2.86	21.96±2.66	21.68±2.28	P>0.05
	Day 7	19±0.41	17.98±0.13	17.96±0.36	17.04±1.55	P>0.05

	Day 14	18.08±0.38	17.82±0.36	17.4±0.82	18.56±0.38	P>0.05
	P-Value	NS	NS	NS	NS	
MCHC	Day 1	35.04 ^{##} ±0.46	27.78±6.57	38.76 ^{###} ±1.51	39.32 ^{##} ±4.19	P>0.05
	Day 7	31.76 [#] ±0.6	29.72±0.32	29.86 [#] ±0.64	26.88 [#] ±2.58	P>0.05
	Day 14	35.46 ^{##} ±0.47	35.08±0.27	33.32 ^{##} ±0.77	34.6 [#] ±0.96	P>0.05
	P-Value	**P<0.01	NS	**P<0.01	*P<0.05	
MCV	Day 1	56.74 ^{##} ±0.66	53.3±7.33	44.98 [#] ±6.46	55.21 [#] ±0.88	P>0.05
	Day 7	59.86 ^{##} ±1.24	60.52±0.73	60.74 ^{##} ±2.02	63.58 ^{##} ±0.85	P>0.05
	Day 14	52.24 [#] ±1.75	50.72±0.8	52.16 [#] ±1.99	53.82 [#] ±1.93	P>0.05
	P-Value	**P<0.01	NS	*P<0.05	**P<0.01	
MO%	Day 1	8.52±2.54	8.04±2.46	9.62±2.75	10.38±2.86	P>0.05
	Day 7	5.76±2.75	4.76±0.67	12.72±3.37	7.98±3.25	P>0.05
	Day 14	9.6±5.08	9.24±5.43	5.7±4.19	6.86±4.16	P>0.05
	P-Value	NS	NS	NS	NS	
MPV	Day 1	5.6 ^{b#} ±0.09	5.68 ^a ±0.26	5.22 ^{b#} ±0.09	5.88 ^a ±0.12	**P<0.01
	Day 7	6.26 ^{##} ±0.1	6.1±0.15	6.48 ^{##} ±0.18	6.1±0.21	P>0.05
	Day 14	5.9 [#] ±0.13	6.28±0.11	6.06 ^{##} ±0.17	6.14±0.12	P>0.05
	P-Value	**P<0.01	NS	**P<0.01	NS	

PCT	Day 1	0.27±0.02	0.32±0.08	0.15±0.03	10.21±9.95	P>0.05	
	Day 7	0.22±0.06	0.39±0.02	0.33±0.07	0.34±0.06	P>0.05	
	Day 14	0.35±0.02	0.3±0.04	0.26±0.05	0.26±0.05	P>0.05	
	P-Value	NS	NS	NS	NS		
PDW	Day 1	36.76±0.98	38.78±1.88	39.86±4.04	37.14±1.32	P>0.05	
	Day 7	41.84±3.92	34.88±0.33	39.82±2.85	38.56±1.61	P>0.05	
	Day 14	35.42±2.06	41.22±2.73	38.8±3.08	39.34±1.98	P>0.05	
	P-Value	NS	NS	NS	NS		
PLT (x10³)	Day 1	479±42.56	541±106.2	284±68.84	437±69.82	P>0.05	
	Day 7	353±93.49	520±130.3	513±117	569±101.1	P>0.05	
	Day 14	478±85.7	476±70.63	431±76.62	420±71.34	P>0.05	
	P-Value	NS	NS	NS	**P<0.01		
RBC	Day 1	6.8±0.56	7.26±0.36	10.58±2.24	6.46±0.54	6.25±0.99	P>0.05
	Day 7	7.62±0.61	8.43±0.19	8.32±0.38	7.7±0.35	9.03±1.15	P>0.05
	Day 14	7.97±0.22	9.54±1.71	8.02±0.45	6.66±0.89	7.13±0.7	P>0.05
	P-Value	NS	NS	NS	NS	NS	
RDW	Day 1	17.28±0.17	16.98±0.2	23.14±5.5	17.14±0.29	17.3±0.29	P>0.05

	Day 7	17.46 ^b ±0.35	17.2 ^b ±0.29	17.94 ^b ±0.47	18.68 ^b ±0.73	19.76 ^a ±0.96	*P<0.05
	Day 14	18.58±0.64	21.6±4.23	17.64±0.34	18.02±0.35	17.74±0.66	P>0.05
	P-Value	NS	NS	NS	NS	NS	
WBC	Day 1	9.2 ^a ±2.77	7.54 ^a ±1.36	5.88 ^a ±1.1	7.06 ^a ±1.49	0.93 ^{##} ^b ±0.3	*P<0.05
	Day 7	8.92±3.08	8.16±2.91	9.68±1.08	11.9±2.75	6.06 [#] ±1.05	P>0.05
	Day 14	8.82±0.37	10.14±2.45	37.08±28.03	8.82±2.12	7.76 [#] ±1.88	P>0.05
	P-Value	NS	NS	NS	NS	**P<0.01	

WBC(**x10³/ul**)-white blood cell, HGB(**g/dl**)-haemoglobin, RBC(**x10⁶/ul**)- red blood cells count, MCV(fl)- mean corpuscular volume, MCH(pg)-mean corpuscular haemoglobin, MCHC(**g/dl**)- mean corpuscular haemoglobin concentration, RDW(%)- red density width, GR(%)- granulocytes count, HCT(%)-hematocrit, PLT(**x10³/ul**)-platelets, LY(%)- lymphocytes, MPV(fl)-mean platelet volume, MO(%)- monocytes, PCT(%)-platelet crit, PDW(%)-platelet density width

DW-Distilled water; All values are expressed as Means±SEM of five animals in each group

Note: P>0.05- Not Significant, *P<0.05-Significan **P<0.01-Significant

The different number of # (in columns) shows a significant difference across the sampled means across the days.

Different superscript letters (in rows) show that the mean is significant from others;

NS –No Significant difference in days across the columns.

363 **Effect of *A. barbadensis* on Lipid parameters**

364 ***High-density lipoprotein cholesterol (HDL-C).***

365 The extract at the administered doses had no significant effect ($P>0.05$) on HDL-C, as
 366 shown in Table 2 on days 1, 7 and 14. The extract at 100 mg/kg had the highest effect on HDL,
 367 with 221.92 ± 58.1 mg/dl concentrations on day 14.

368

369 **Table 2.** Effect of ethanol extract of *A. barbadensis* on high-density lipoprotein cholesterol
 370 (HDL-C) concentrations (mg/dl) of male rats.

Groups	Day 1	Day 7	Day 14	
Control(DW)	52.18 [#] ±14.53	118.45 [#] ±30.63	133.72 ^{##} ±9.55	*P<0.05
100mg/kg	54.21 [#] ±23.96	126.41 [#] ±33.7	221.92 ^{##} ±58.1	*P<0.05
200mg/kg	54.21 [#] ±23.96	94.1 [#] ±38.51	211.15 ^{##} ±18.94	**P<0.01
400mg/kg	26.02 [#] ±16.1	160.94 ^{##} ±4.72	128.75 ^{##} ±18.33	**P<0.01
P-value	P>0.05	P>0.05	P>0.05	

371 DW-Distilled water; All values are expressed as Means±SEM of five animals in each group
 372 Note: **P<0.01-Highly significant, *P<0.05-Significant, P>0.05-Not Significant
 373 Same # (in rows) shows no significant difference across the sampled means across the days. NS –
 374 No Significant difference in days across the rows
 375

376 ***Low-density lipoprotein cholesterol (LDL-C)***

377 Table 3 revealed the effect of the extract on LDL-C. The administered doses of the extract
 378 to the male rats had no significant effect ($P>0.05$) on LDL-C, on days 1 and 14 (Table 3). The
 379 highest concentration of LDL-C observed at the 200 mg/kg dose level was 150.05 ± 15.19 mg/dl.

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384 **Table 3.** Effect of ethanol extract of *A. barbadensis* on low-density lipoprotein cholesterol
 385 (LDL-C) concentrations (mg/dl) of male rats.

Groups	Day 1	Day 7	Day 14	P-Value
Control(DW)	109.62±15.99	105.15 ^a ±7.3	65.48±18.55	NS
100mg/kg	113.21±32.86	29.39 ^b ±12.11	99.43±57.16	NS
200mg/kg	54.83±22.85	78.55 ^a ±23.66	150.05±15.19	NS
400mg/kg	47.69 [#] ±15.59	119.3 ^{a##} ±11.64	84.64 [#] ±18.56	*P<0.05
P-value	P>0.05	**P<0.05	P>0.05	

386 DW-Distilled water; All values are expressed as Means±SEM of five animals in each group
 387 Note: **P<0.01-Highly significant, *P<0.05-Significant, P>0.05-Not Significant
 388 Different superscript letters (in columns) show that the mean is significant from others.
 389 Same # (in rows) shows no significant difference across the sampled means across the days. NS –
 390 No Significant difference in days across the rows

391
 392 **Triglyceride (TRIG)**

393 The effect of the extract on TRIG concentrations of all same dose groups at different days
 394 was non- significant. The administered doses of the extract to the male rats had no significant
 395 effect (P>0.05) on TRIG on days 1 and 14 (Table 4).

396 **Table 4.** Effect of ethanol extract of *A. barbadensis* on triglyceride concentrations (mg/dl) of
 397 male rats.

Groups	Day 1	Day 7	Day 14	P-Value
Control(DW)	122.99±36.17	66.86 ^b ±24.23	136.42±33.14	NS
100mg/kg	116.62±32.03	253.22 ^a ±40.91	215.09±36.97	NS
200mg/kg	77.69±23.53	99.09 ^b ±20.14	121.38±36.9	NS
400mg/kg	132.05±36.64	121.65 ^b ±47.77	131.04±39.96	NS
P-value	P>0.05	**P<0.01	P>0.05	

398 DW-Distilled water; All values are expressed as Means±SEM of five animals in each group
 399 Note: **P<0.01-Highly significant, *P<0.05-Significant, P>0.05-Not Significant
 400 Different superscript letters (in columns) show that the mean is significant from others.
 401

402 *NS –No Significant difference in days across the rows*

403

404 **Total cholesterol concentrations**

405 A significant decrease in total cholesterol concentrations as the doses increased on days 1, 7
406 and 14 were observed (Table 5). On day 1, there was no significant difference in the effect of all
407 the tested doses on cholesterol but were significantly different ($P < 0.01$) from the control
408 (distilled water).

409 **Table 5.** Effect of ethanol extract of *A. barbadensis* on total cholesterol concentrations (mg/dl)
410 of male rats.

Groups	Day 1	Day 7	Day 14	P-Value
DW	184 ^{a###} ±19.03	62.26 ^{#b} ±4.4	71.96 ^{b#} ±14.38	**P<0.01
100mg/kg	113.89 ^b ±42.53	186.04 ^a ±12.15	145.39 ^a ±26.47	NS
200mg/kg	99.83 ^b ±28.35	84.78 ^b ±16.62	63.52 ^b ±12.82	NS
400mg/kg	64.28 ^b ±18.5	67.02 ^b ±5.83	46.73 ^b ±12.13	NS
P-value	**P<0.01	**P<0.01	**P<0.01	

411 DW-Distilled water; All values are expressed as Means±SEM of five animals in each group
412 *Note: **P<0.01-Highly significant, *P<0.05-Significant, P>0.05-Not Significant*
413 *Different superscript letters (in columns) show that the mean is significant from others.*
414 *Same # (in rows) shows no significant difference across the sampled means across the days. NS –*
415 *No Significant difference in days across the rows*

416

417 **DISCUSSION**

418 The haematological and lipid profiles of male Wistar rats were evaluated following the
419 assessment of functional biochemical indices or “markers.” Reports show that a significant
420 change in any organ function's biochemical indices will impair the normal function of the organ
421 (Afolayan and Yakubu, 2009). Therefore, haematological and lipid parameters are essentially

422 necessary tools in clinical diagnosis and toxicological studies like serum enzyme levels (Ashafa
423 et al. 2009).

424 Haematological parameters have been used to unravel the magnitude of the harmful effect of
425 foreign substances such as plant extracts on an animal's blood constituents. Also, haematological
426 parameters had been used to explain blood-relating functions of chemical compounds, including
427 extracts of plants (Yakubu et al. 2007). The non-significant ($p>0.05$) effect following the oral
428 administration of the extract on RBC and indices relating to it (HGB, PCT, MCV, MCH, and
429 MCHC) (Table 1) indicates that there was no destruction of matured RBC's and erythropoiesis
430 was not changed. Furthermore, the blood's oxygen-carrying capacity was unhindered because of
431 the non-significant effect of the extract on RBC and HGB (de Gruchy, 1976). The blood
432 diagnostic parameters (MCV, MCH and MCHC) of anaemia (Coles, 1986), as displayed in Table
433 1, were not affected, suggesting an unremarkable effect on the average size of RBC (microcytes)
434 and haemoglobin as well as the weight per RBC. These findings imply that the extract cannot
435 induce anaemia within 14 days of administration. The WBC and all indices relating to it (GR,
436 LY, HCT and MO) were not altered. It implies that the ability of the animal to eliminate
437 infection was not affected. It also suggests that there was an unremarkable stimulation of the
438 immune system. The platelets -blood cells involved in coagulation (Williams and Levine, 1982)
439 were not significantly ($p>0.05$) altered. It indicates that the extract did not adversely affect the
440 platelets' size, number, and function. The extract had no significant localized systemic toxicity,
441 affecting the WBC's normal functioning and related indices. These findings were dissimilar to
442 previous reports by Adebayo et al. (2005) on the ethanolic extract of *Bougainvillea spectabilis*;
443 Yakubu et al. (2007), and Yakubu and Afolayan (2009) on the aqueous extracts of *Fadogia*
444 *argrestis* stem and *Bulbine natalensis* stem.

445 The concentrations of major lipids such as cholesterol, high-density lipoprotein cholesterol
446 (HDL-C), low-density lipoprotein cholesterol (LDL-C) and triglycerides, when altered, can give
447 vital information on the metabolism of lipids. These alterations can also give useful information
448 on the predisposition of the heart to atherosclerosis and other associated cardiovascular diseases
449 (Yakubu et al. 2008). The administration of the extract significantly ($p<0.01$) decreased
450 cholesterol on day 1 but significantly ($p<0.01$) increased it on days 7 and 14 at 100 mg/kg when
451 compared to control (Table 5). This may be due to impairment in cholesterol biosynthesis on day
452 1 and non-impairment in cholesterol biosynthesis on days 7 and 14. This increase in cholesterol
453 can be due to an increase in acetyl CoA concentration as a key substrate in the biosynthesis of
454 cholesterol (Range et al. 1995). However, Treasure et al. (1995) reported that high blood
455 cholesterol concentration is one of the vital risk factors for cardiovascular disease, suggesting
456 that such an increase may not be beneficial to the animals as it may enhance atherosclerosis and
457 hypertension (Enas, 1999). The non-significant ($p>0.05$) effect of the extract on HDL - C (Table
458 2) is an implication that the anti-atherogenic property of HDL-C was not affected. LDL-Cs form
459 plaque that clots the arteries resulting in atherosclerosis. LDL-Cs are primary carriers of
460 cholesterol that build up in the arteries' walls supplying blood to the heart and brain (Ashafa et
461 al. 2009). The non-significant ($p>0.05$) effect of the extract on the lipid parameter-LDL-C (Table
462 3) shows that the extract may not predispose the heart to atherosclerosis. Again, the non-
463 significant ($p>0.05$) effect of the extract on triglyceride (Table 4) implies that lipolysis was not
464 enhanced, which indicates a non-depletion in the storage of fatty acids. It can be inferred that the
465 ethanol extract of *A. barbadensis* may not predispose the male animals to atherosclerosis and
466 other associated coronary heart diseases despite isolated significant alterations in LDL- C,

467 triacylglycerol and cholesterol on day 7 at 100 mg/kg. This increase was alleviated by the non-
468 significant increase in HDL – C ('good cholesterol').

469 CONCLUSIONS

470 The toxicological study shows that the extract was comparatively safe for consumption at the
471 administered doses. There was no significant harmful consequence on the haematological
472 parameters of the male rats. In contrast, there were isolated alterations in the evaluated lipid
473 parameters. Additionally, no significant localized systemic toxicity was noticed. Still, caution is
474 needed when using the extract of *A. barbadensis* root for oral remedies following its potential
475 selective ability to alter specific lipid parameters in male rats. Further studies on the effect of
476 chronic administration of the root of *A. barbadensis* are recommended.

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