

ABSTRACT

The study investigated the adjuvant effect of anxiolytic and anti-inflammatory co-treatment on Isoniazid- induced seizure. A total of twenty (20) healthy adult Wistar rats average weight 210kg were used and divided into five (5) groups of four(4) rats per a group. The animals in groups B, C, D, E were induced seizures by 300mg/kg isoniazid single dose o.p, while animals in group A served as control group and were given 0.1ml of normal saline (placebo). Group B was the positive seizure group untreated while animals in group C were treated with 5mg/kg diazepam and group D were treated with 50mg/kg hydrocortisone, those in group E were treated with combination of 5mg/kg diazepam and 50mg/kg hydrocortisone for 5days respectively. Sections of the hippocampus were immunohistochemical staine for astrocyte expression using GFAP marker. The seizure group exhibited positive immunoreactivity to GFAP with astrogliosis. Hydrocortisone and diazepam groups revealed restored normal astrocytes expression. The combined therapy provided a synergist effect depicted as retraction of most astrocytes. Put together the combined anxiolytic and anti inflammatory should be considered in seizure management.

Keywords: Anxiolytic, Anti Inflammatory, Seizure, Astrocytes, Steroids

INTRODUCTION

Status epilepticus is a life-threatening neurological emergency with high mortality and common in the management of tuberculosis infection (Pitkanen et al., 2005). Tuberculosis (TB) is an airborne disease caused by *Mycobacterium tuberculosis* (Mtb) that usually affects the lungs (Nicole, 2015). Mtb infection is acquired by inhalation of infectious aerosol particles released from close contacts (Cruz-Knight and Blake-Gumbs, 2013; Berry et al., 2013; Nicole, 2015). Although, many inherent factors interplay in the surveillance of M.tb, including the difficulty in obtaining the right drug treatment, drug resistance and comorbidity with infection, anxiety, inflammation and seizure (Toossi, 2000; Smitha and Jordi, 2011; Grab et al., 2019). The seizure episode is often due to the secondary effect of drug treatment particularly with Isoniazid.

Isoniazid (INH) is one of the potent anti mycobacterial agent that inhibits cell wall synthesis of *Mycobacterium tuberculosis* (M.tb) and used in both therapeutic and prophylactic regimens of tuberculosis (Toossi, 2000; Smitha and Jordi, 2011; Blise et al., 2017; Grab et al., 2019). Resistance to INH is common and has been linked to risk of acute neurotoxicity with single high therapeutic dose, which manifests as neurological side effects, including peripheral neuritis, dizziness, and insomnia and seizures (Fekit et al., 2011; Cruz-Knight and Blake-Gumbs, 2013; Nicole, 2015). The important mechanism is a deficiency of gamma-aminobutyric acid (GABA) and pyridoxine induced by INH leading to reduced production of GABA, as it is usually a product of pyridoxine-dependent decarboxylation reaction (Finbarrs-Bello et al., 2019). The hallmark of GABA deficiency manifest as seizures which can be managed with the use of benzodiazepines, and pyridoxine in humans and rodents, this helps with rapid restoration of GABA stores and resolution of the seizure (Yasudishi et al., 2005; Finbarrs-Bello et al., 2019).

Anxiety disorders represent another common psychiatric co-morbidity in patients with epilepsy, affecting prognosis and quality of life (Kroenke et al., 2007). The associated sleep deprivation increases the risk of seizures attacks as well (Mato et al., 2011). Anxiolytics, or anti-anxiety drugs are recommended to prevent and treat anxiety, which targets key chemical messengers in the brain such as dopamine, serotonin, and GABA, to help decrease abnormal excitability (Beldum, 2008; Durant et al., 2010; Mula, 2016). One of the frequently prescribed anxiolytic (benzodiazepine) diazepam suppresses generalized seizures and effectively stops absence, infantile, and other myoclonic seizures and even restores normal sleep patterns (Ravindran et al., 2010; Mula, 2016). They are the drug of choice to arrest status epilepticus or all kinds of eclamptic convulsions (Ravindran et al., 2010).

In addition, the core of immune response to every infection is inflammation; there are some reports on the development of inflammatory reactions during seizure attacks, including increased release of chemokines, pro-inflammatory cytokines and prostaglandins in the brain of rodents (Vezzani et al., 2002; Vezzani and Tiziana, 2005). It is well established that inflammatory mediators also are produced by brain parenchymal cells (microglia, astrocytes, and neurons) and by cells of the BBB and choroid plexus (Vezzani, 2012). Also, there is evidence that inflammation, has significant effects on blood brain barrier (BBB) integrity, affects normal brain function and contributes to the pathophysiology of seizures (Marchi et al. 2007). Insight into the inflammatory response supports the application of anti-inflammatory agents as adjuvants in the treatment of tuberculosis (Toossi, 2000; March et al., 2011; Zumla et al., 2014). As such corticosteroids treatment have been proven to be beneficial to survival of TB patient even with seizure for example hydrocortisone (Senderovits and Viskum ,1994; Grosso et al., 2008; Schutz et al., 2018; Grab et al., 2019).

These commonalities highlight the possibility that both anti inflammatory and anxiolytics or even the combination would play a potential role in treatment strategies for seizure. In the study we targeted brain resident innate immune cell the astrocytes which are involved in promoting neuronal survival and drug metabolism in the brain, using an animal model of chemical induced status epilepticus seizure that can be seen present in an infection like tuberculosis.

METHODS

Drugs

Isoniazid PubChem CID; 3767 (Seizure inducing drug), Diazepam PubChem CID; 3016 (Anxiolytic agent), Hydrocortisone (Solu-cortef) PubChem CID; 5754(Anti-inflammatory drug) were procured from the registered pharmacist at Enugu state.

Animals

The experiments were performed on 20 inbred adult wistar rats (11 weeks old) weighing 210 ± 20 g and age-matched. The rats were housed in the animal facility of the animal house of ESUT College of Medicine. Enugu Nigeria. The rats were housed (4 per cage) under standardized conditions (25°C, 40–50% humidity; 12/12-h light/dark cycle, with light on at 6:00 a.m.) and habituated for a week before experiments. Food and water were available ad libitum throughout the study.except. The experimental procedures and techniques used in the study were in accordance with accepted principles for laboratory animal use and care by NIH, 1985 and EU directive of 1989:86/609/EEC. The protocol used was reviewed and approved by the Research Ethics Committee of Faculty of Basic Medical Science (REC-FBMS), Enugu State University of Science and Technology.

Experimental Design

Group 1: 0.1ml normal saline

Group 2: 300mg/kg/bw Isoniazid (single dose, p.o)

Group 3: 300mg/kg/bw Isoniazid +5mg/kg/bw Diazepam/5days

Group 4: 300mg/kg/bw Isoniazid +50mg/kg/bw Hydrocortisone/5days

Group5:300mg/kg/bw Isoniazid+ 5mg/kg/bw Diazepam +50mg/kg/bw Hydrocortisone/5days

Tissue processing and Immunohistochemistry

At the sixth day, the animals were deeply anesthetized with ether anesthesia and sacrificed except the group 2 that was sacrificed on the day 1 post- induction of seizure. Brains were fixed in 10% neutral formal saline solution and embedded in paraffin. Sectioned of 10um thickness with of each representative group were deparaffinized and further processed. Immunoperoxidase was used to label astrocytes using glial fibrillary acidic protein (GFAP) (Novocastra, LEICA Germany) as a marker. Endogenous peroxidase activity was blocked with pre-incubation in 0.3% H₂O₂. After washing, the sections were pre-incubated for 1 h at room temperature in the appropriate normal serum before incubation in primary antibodies overnight at 4°C. The sections were then rinsed and incubated in secondary antibodies at 1:200 dilution for 2 h at room temperature, and then reacted in avidin biotin complex solution (Novocastra, LEICA Germany) for 1.5 h using 3,3'-diaminobenzidine (DAB) as chromogen. The sections were then mounted on slides, dried, dehydrated, cleared and cover slipped with DPX. The slides were interpreted and photographed. Star shaped cells with specific dark brown colours in the cytoplasm or nuclei depending on the antigenic sites are considered to be positive. The haematoxylin stained cells without this form are scored negative. Nonspecific binding/brown artifacts on cells and connective tissue were disregarded.

RESULTS

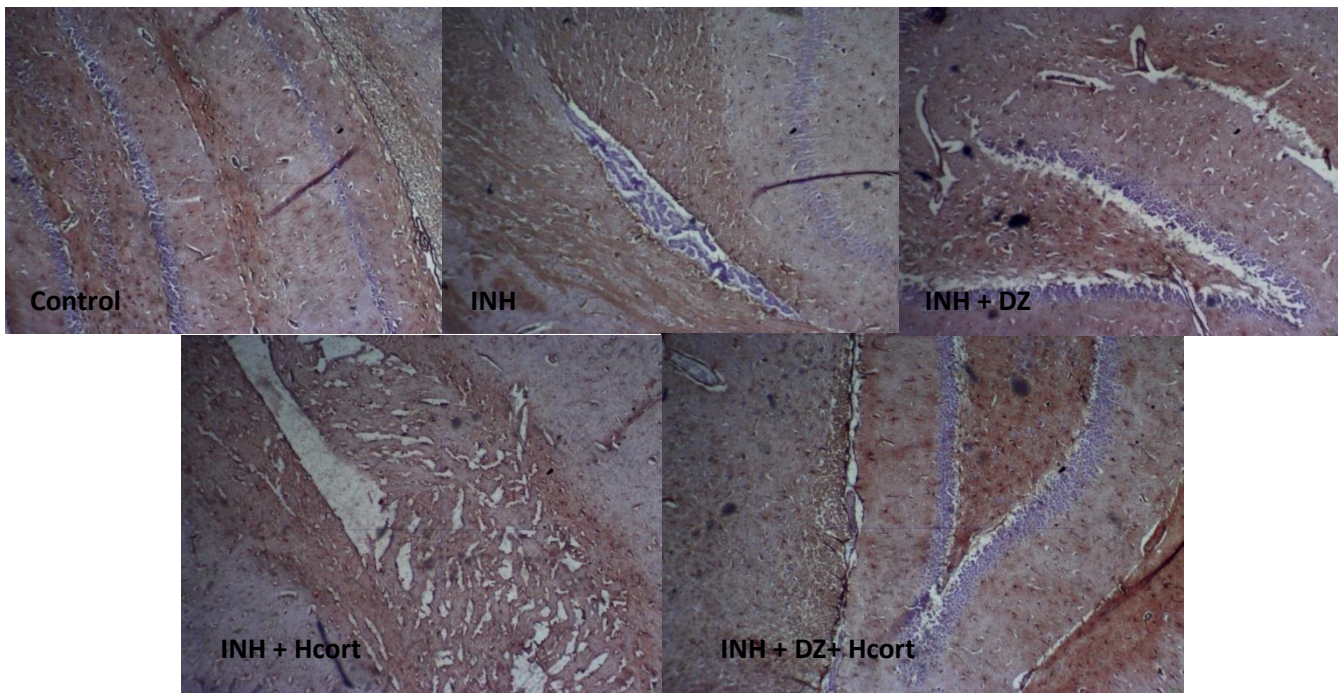


Figure 1: Sections of hippocampus of control (Normal astrocytes). Isoniazid (INH) shows reactive astrocytes. Anxiolytic treated group (INH +DZ) Mild expression of reactive astrocytes. Anti-inflammatory treated group (INH + Hcort) mild astrocytes retraction. Co-treatment (INH + DZ+ Hcort) severe retraction of astrocytes. GFAP. x200.

Astrocytes are morphologically star shaped glial cells found in the brain and spinal cord. Astrocytes support neurons, endothelial cells of blood-brain-barrier, provide nutrients and conducive environment for neuronal survivals (Nag, 2011). Astrocytes have been implicated to the maintenance of the extracellular ion balance, repair and scarring process of the CNS; particularly following injuries and disease conditions (Bylicky et al., 2018). This glial cell exhibits morphological diversity, plasticity and disease related deficits by changing their phenotypes and population (Zhou et al., 2019). In the present study, astrocyte was demonstrated by Glia fibrillary acidic protein (GFAP) which showed dark star shaped astrocytes with various processes. The hallmark of astrocytic expression was seen in the seizure model (Fig 1: INH), marked by astrocytes that become reactive in expression with prominent processes. This is consistent with astrocytic morphological phenotypes in neurotoxicity (Finbarrs-Bello et al., 2016). It is a common occurrence to observe astrocytes in

negative responses through their hyperreactivity and glial scar formation in excitotoxic and/or mechanical injuries (Becerra-Calixto et al., 2017).

Although, reactive astrocytosis may tend to have biphasic characteristic, one for cell death and one for pro-neuroprotection probably depending on the context (Becerra-Calixto et al., 2017). The morphological and physiological changes that astrocytes undergo in response to pathology is most often characterized by an up-regulation of the astrocytic intermediate filament glial fibrillary acidic protein (GFAP) and cellular hypertrophy, which may or may not be associated with cell proliferation (Rivera et al., 2019). This suggests that isoniazid induced neuronal damage which resulted into the activation of astrocytes. Conversely, it could also be attributed to isoniazid induced reduction in GABAergic neurotransmission of which astrocytes play pivotal role GABA synthesis and release at synaptic terminals (Carta and Luca, 2008).

Anxiolytic treatment resulted in the retraction of astrocytes which is a reversal of the seizure effect. By implication diazepam facilitates the action of GABA, decreased the neuronal excitability and necessitates astrocytes retraction. Similarly, anti-inflammatory drug hydrocortisone was used to evaluate the possible modulatory effect of anti-inflammatory agent as adjuvant in seizure management. Hydrocortisone, exhibits a better promising effect by restoring astrocytes to relative normal morphology. Grosso et al., (2008) reported that hydrocortisone could be useful in the treatment of severely drug resistant, childhood epilepsies. Furthermore, a previous study demonstrates corticosteroids steroid which includes hydrocortisone are safe and efficient for treatment of epilepsy, particularly short therapy (Buzatu et al., 2009). This was evident in this study whereby hydrocortisone was able to ameliorate the astrogliosis induced by isoniazid.

The combination of anxiolytic and anti-inflammatory treatments reveals marked positive immunoreactivity and retraction of astrocytes as shown by their morphology. The above

result demonstrated the beneficial effect of the combined therapy to enhance the management of status epilepticus. We opined that diazepam enhanced GABA release in the process while the hydrocortisone might have acted via the corticosteroid hormone receptor and/or exerted adjunct effect in the course of the treatment.

Conclusion

The combination of anxiolytic and anti-inflammatory therapy could be a promising treatment for seizure. Thus, we advocate the combination of both therapies in the management of seizure patients to better treatment outcome.

Conflicts of Interest

Authors have declared that they have no competing interest

Acknowledgment

We thank the staff of the histology laboratory for providing the technical assistance.

REFERENCE

Baldwin DS (2008). Room for improvement in the pharmacological treatment of anxiety disorders. *Current Pharmaceutical Design*. 14:4382–3491.

Becerra-Calixto A, Cardona-Gómez GP (2017). The role of astrocytes in neuroprotection after brain stroke: potential in cell therapy. *Front Mol Neurosci*.10: 1-12.

Berry M.P.R, Blankley S, Graham C.M, Bloom C.I, O'Garra A (2013). Systems approaches to studying the immune response in tuberculosis. *Curr Opin Immunol*. 25 (5):579-587.

Bhise S.B (2017). Isoniazid Toxicity. *J Drug Des Res*. 1060; 4(7):2-8.

Buzatu M, Bulteau C, Altuzarra C, Dulac O, Van Bogaert P.(2009). Corticosteroids as treatment of epileptic syndromes with continuous spike waves during slow-wave sleep. *Epilepsia*. 50(7):68-72.

Bylicky MA, Mueller GP, Day RM (2018). Mechanisms of Endogenous Neuroprotective Effects of Astrocytes in Brain Injury. *Oxid Med Cell Longev*. 2018;1-16.

Carta Mario, Luca Murru (2008). Isoniazid-induced reduction in GABAergic neurotransmission alters the function of the cerebellar cortical circuit. *Neuroscience*. 154(2):710-9.

Cruz-Knight W, Blake-Gumbs L (2013). Tuberculosis: an overview *Prim Care*.40 (3):743-756.

Durant C, Christmas D, Nutt D (2010). The pharmacology of anxiety. *Curr Top Behav Neurosci*.2:303-30.

Fekih L, Boussoffara L, Fenniche S, Abdelghaffar H, Megdiche ML (2011). Neuropsychiatric side effects of antituberculosis agents. *Rev Med Liege*. 66(2):82-5.

Gräb J, Suárez I, van Gumpel E, Winter S, Schreiber F, Esser A et al (2019). Corticosteroids inhibit Mycobacterium tuberculosis-induced necrotic host cell death by abrogating mitochondrial membrane permeability transition. *Nature communications*.10(688):1-14.

Grosso S, Farnetani M, Mostardini R, Cordeli D, Berardi R, Balestri P (2008). Comparative study of hydrocortisone versus deflazacort in drug-resistant epilepsy of childhood. *EpilepsyReps*. 81(1):80-85.

Kälviäinen R, Eriksson K, Parviainen I(2005). Refractory generalised convulsive status epilepticus: a guide to treatment. CNS Drugs.19 (9): 759-68.

Kroenke K, Spitzer RL, Williams JB, Monahan PO, Lowe B (2007). Anxiety disorders in primary care: Prevalence, impairment, co-morbidity and detection. *Ann Intern Med*.146:317–25.

Marchi N, Angelov L, Masaryk T, Fazio V, Granata T, et al (2007). Seizure-promoting effect of blood-brain barrier disruption. *Epilepsia*.48: 732–742.

Marchi N, Granata T, Freri E, Ciusani E, Ragona F, Puvenna V, et al (2011). Efficacy of Anti-Inflammatory Therapy in a Model of Acute Seizures and in a Population of Pediatric Drug Resistant Epileptics. *PLoS ONE*.6(3): e18200;1-10.

Matos G., Tufik S., Scorza F. A., Cavaleiro E. A., Andersen M. L (2011). Sleep, epilepsy and translational research: what can we learn from the laboratory bench. *Prog. Neurobiol.* 95, 396–405.

Mula M (2016). Using anxiolytics in epilepsy: neurobiological, neuropharmacological and clinical aspects. *Epileptic Disord.*18(3):217-27.

Nag. S (2011). Morphology and properties of astrocytes. *Methods Mol Boil.* 686:69-100.

Nicole F (2015). Tuberculosis: A disease without boundaries. *Tuberculosis.* 95(1):527-531

Olubunmi A.O. (2006). Seizure in Nigeria: A review of etiology, epidemiology and management. *Journal of Postgraduate Medicine*, 8: 27–51.

Pitkänen A, Kharatishvili I, Narkilahti S, Lukasiuk K, Nissinen JA (2005). Administration of diazepam during status epilepticus reduces development and severity of epilepsy in rat. *Epilepsy Res.* 63(1):27-42.

Ravindran LN., Stein MB (2010). The pharmacologic treatment of anxiety disorders:a review of progress. *J Cli Psychiatry.* 71(7):839-854.

Rivera, A.D., Butt, A.M (2019). Astrocytes are direct cellular targets of lithium treatment: novel roles for lysyl oxidase and peroxisome-proliferator activated receptor- γ as astroglial targets of lithium. *Translational Psychiatry* 9(211):1-14.

Schutz C, Davis AG, Sosen B, Lair P, Ntsekhe M, Harley YX, Wilkinson RJ (2018). Corticosteroids as an adjunct to tuberculosis therapy. *Expert Review of Respiratory Medicine.*12(10):881-891.

Sendevoulz T, Viskum K(1994). Corticosteroids and tuberculosis . *Respir Med.*88(8);561-565.

Smitha J S, Jordi B T (2011). *Mycobacterium tuberculosis* infection and inflammation, what is beneficial for the host and for the bacterium. *Front microbial.*2(2):1-16.

Sofroniew, Michael V (2009). Molecular dissection of reactive astrogliosis and glial scar formation. *Trends in Neurosciences.* 32(12):638-47.

Toossi Z (2000). The inflammatory response to *Mycobacterium tuberculosis* infection. *Arch immunol. Ther. Exp(warsz).*48(6):513-519.

Vezzani A, Tiziana G (2005).Critical Review; Brain Inflammation in Epilepsy: Experimental and Clinical Evidence. *Epilepsia.*46(11):1724–1743.

Vezzani A (2012). Inflammation and epilepsy. *HandbClin Neurol.*107:163–175.

Vezzani A. Moneta D. Richichi C. Aliprandi M. Burrows SJ. Ravizza T (2002). Functional role of inflammatory cytokines and anti-inflammatory molecules in seizures and epileptogenesis. *Epilepsia.*43(5):30-35.

Yasuloshi I, Masashi N, Masato Y (2005). Effect of isoniazid on the pharmacodynamics of Cefazolin induced seizures in rats. *Drug Metab Pharmacokinet.* 20(2):117-120.

Yu-Chen Chen,Ning-Xin Ma, Zi-Fei Pei, Kathryn Lee,Gregory J. Quirk,Gong Chen. A(2020). NeuroD1 AAV-Based Gene Therapy for Functional Brain Repair after Ischemic Injury through In Vivo Astrocyte-to-Neuron Conversion. *molecular therapy.*28(1): 217-234,

Zhou B, Zuo YX, Jiang RT (2019). Astrocytes morphology: diversity, plasticity and role in neurological disease. *CNS Neurosci.Ther.*25(6):665-673.

Zumla A, Rao M, Parida S K, Keshavjee S, Cassell G, Wallis R, et al (2015). Inflammation and tuberculosis: host-directed therapies. *Journal of Internal Medicine.*277; 373-387.